

## Cytotoxic Principles from *Ventilago leiocarpa*

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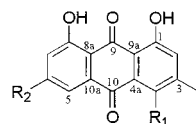
Three new anthraquinones, islandicin 4-methyl ether (**1**), 1,2,6-trihydroxy-7,8-dimethoxy-3-methylanthraquinone (**2**), and 2-hydroxyemodin 1-methyl ether (**3**) as well as two known triterpenoids [taraxerol (**4**), lupeol (**5**)], six anthraquinones [chrysophanol (**6**), islandicin (**8**), parietin (**9**), emodin (**10**), catenarin (**11**), skyrin (**15**)], a 2,3-dihydroflavonol [(+)-aromadendrin (**12**)], two benzisochromanquinones [ventiloquinone K (**13**) and ventiloquinone I (**14**)], and stigmasterol (**7**) were isolated from *Ventilago leiocarpa*. The cytotoxicity of these compounds to various tumor cell lines was evaluated, and compound **15** significantly suppressed growth of HeLa, Vero, K562, Raji, Wish, and Calu-1 tumor cell lines. With the exception of K562 cells, the proliferation of other tumor cell lines was inhibited by compounds **3** and **10**.

*Ventilago leiocarpa* Benth. belongs to the Rhamnaceae family, which is a scandent glabrous shrub growing throughout the thickets at low and medium altitudes in Taiwan.<sup>1</sup> It is used in Chinese folk medicine as an analgesic and for the treatment of rheumatism.<sup>2</sup> Previous chemical studies<sup>3–7</sup> of the genus *Ventilago* have shown the presence of a variety of anthraquinones, naphthoquinones, quinones, and benzisochromanquinones. In the course of our search for biologically active substances in nature, we found that the crude extract from the stems of *V. leiocarpa* possessed in vitro cytotoxicity to cancer cells. Since the bioactivity and chemical constituents of *V. leiocarpa* have not been studied, we investigated the chemical constituents of dried stems of this plant.

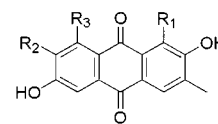
The ethanolic extract of the stems of *V. leiocarpa* was fractionated by solvent partition and separated by column chromatography to yield anthraquinones, benzisochromanquinones, 2,3-dihydroflavonol, and phytosterol. The structures of **4–15** have been established as taraxerol (**4**),<sup>8</sup> leupol (**5**),<sup>9</sup> chrysophanol (**6**),<sup>10</sup> stigmasterol (**7**), islandicin (**8**),<sup>11</sup> parietin (**9**),<sup>12</sup> emodin (**10**),<sup>13</sup> catenarin (**11**), (+)-aromadendrin (**12**),<sup>14,15</sup> ventiloquinone K (**13**),<sup>3</sup> ventiloquinone I (**14**),<sup>3</sup> and skyrin (**15**),<sup>16</sup> on the basis of spectral analyses and by comparison with reported data.

Compound **1** has the molecular formula C<sub>16</sub>H<sub>12</sub>O<sub>5</sub>, which was confirmed by HREIMS. The IR spectrum shows absorption bands 1667 and 1620 cm<sup>-1</sup>, and its UV spectrum (λ<sub>max</sub> 441 nm) suggested a 1,8-dihydroxyanthraquinone structure. The <sup>1</sup>H NMR spectrum shows the presence of an aromatic methyl (δ 2.41), two *peri*-hydroxyl protons (δ 12.06, 12.51), four aromatic protons (one singlet at δ 7.18, and an ABC system at δ 7.27, 7.68, 7.81), and a methoxyl group at δ 3.89. Acetylation of **1** with acetic anhydride/pyridine afforded a diacetate (**1a**). The <sup>1</sup>H NMR data of **1** showed similarities to those of islandicin,<sup>6</sup> except for the presence of a methoxyl group. In the HMBC spectrum, the C-4 signal at δ 153.8 was correlated with resonances at δ 3.89 (OCH<sub>3</sub>-4), 7.18 (H-2), and 2.41 (CH<sub>3</sub>-3), and the signal at δ 126.9 (C-2) correlated with the 1-OH signal (δ 12.51) and CH<sub>3</sub>-3 (δ 2.41), indicating that C-4 was methoxylated. Therefore, compound **1** was identified as the new compound, islandicin 4-methyl ether.

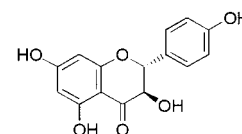
Compound **2** has the molecular formula C<sub>17</sub>H<sub>14</sub>O<sub>7</sub>, which was confirmed by HREIMS. The <sup>1</sup>H NMR spectrum of **2**



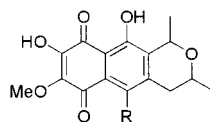
- 1: R<sub>1</sub> = OMe R<sub>2</sub> = H  
 6: R<sub>1</sub> = R<sub>2</sub> = H  
 8: R<sub>1</sub> = OH R<sub>2</sub> = H  
 9: R<sub>1</sub> = OH R<sub>2</sub> = OMe  
 10: R<sub>1</sub> = H R<sub>2</sub> = OH  
 11: R<sub>1</sub> = OH R<sub>2</sub> = OH



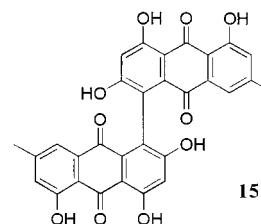
- 2: R<sub>1</sub> = OH R<sub>2</sub> = R<sub>3</sub> = OMe  
 3: R<sub>1</sub> = OMe R<sub>2</sub> = H R<sub>3</sub> = OH



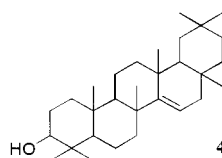
**12**



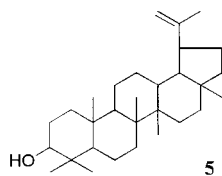
- 13: R = OMe  
 14: R = H



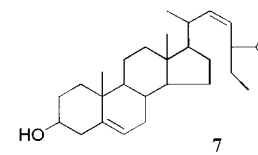
**15**



**4**



**5**



**7**

shows signals for an aromatic methyl (δ 2.35), two methoxyl groups (δ 3.98, 3.99), two isolated aromatic protons (δ 7.55 and 7.57), and three hydroxyl protons (one *peri*-OH at δ 13.32, and two free-OH at δ 8.92, 9.63), which were confirmed by formation of a triacetate derivative (**2a**). The HMBC spectrum revealed the three-bond coupling of C-9a (δ 116.3) to H-4 (δ 7.55) and OH-1 (δ 13.32), C-2 (δ 150.8) to CH<sub>3</sub>-3 (δ 2.35) and H-4 (δ 7.55), C-10 (δ 181.0) to H-4 (δ 7.55) and H-5 (δ 7.57), and C-7 (δ 147.3) to OCH<sub>3</sub>-7 (δ 3.98) and H-5 (δ 7.57), indicating compound **2** possessed a 1,2-dihydroxy-3-methylanthraquinone component. In the NOESY spectrum, no correlations were observed between

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**Table 1.** Inhibitory Activity (%) of Compounds **3**, **10**, and **15** on Various Tumor Cells Growth<sup>a</sup>

compound	dose ( $\mu\text{M}$ )	cell line					
		Calu-1	HeLa	K562	Raji	Vero	Wish
<b>3</b>	100	93.8 $\pm$ 7.8	100 $\pm$ 8.8	34.1 $\pm$ 7.8	100 $\pm$ 6.8	85.9 $\pm$ 2.5	79.3 $\pm$ 5.6
	50	91.7 $\pm$ 5.4	49.9 $\pm$ 7.2	N.D.	100 $\pm$ 7.2	71.1 $\pm$ 3.1	56.0 $\pm$ 3.8
	25	71.0 $\pm$ 3.3	21.4 $\pm$ 2.9	N.D.	72.4 $\pm$ 6.3	42.6 $\pm$ 2.8	-0.30 $\pm$ 3.2
	12.5	60.5 $\pm$ 6.8	1.3 $\pm$ 4.2	N.D.	71.7 $\pm$ 5.0	13.8 $\pm$ 3.2	-5.5 $\pm$ 2.9
	6.25	-4.8 $\pm$ 3.3	-22.3 $\pm$ 3.6	N.D.	63.7 $\pm$ 4.9	9.9 $\pm$ 4.0	-9.5 $\pm$ 5.1
IC <sub>50</sub> ( $\mu\text{M}$ )		21.3 $\pm$ 5.0	50.0 $\pm$ 6.2	>100	<6.25	32.5 $\pm$ 4.5	55.0 $\pm$ 6.4
<b>10</b>	100	100 $\pm$ 7.5	100 $\pm$ 5.3	29.5 $\pm$ 6.5	100 $\pm$ 6.5	69.1 $\pm$ 2.5	80.7 $\pm$ 4.5
	50	100 $\pm$ 6.8	100 $\pm$ 6.1	N.D.	47.2 $\pm$ 4.9	64.4 $\pm$ 4.1	63.3 $\pm$ 6.3
	25	100 $\pm$ 5.3	78.3 $\pm$ 3.8	N.D.	45.8 $\pm$ 6.1	26.7 $\pm$ 3.8	61.9 $\pm$ 4.7
	12.5	60.6 $\pm$ 4.8	41.9 $\pm$ 3.5	N.D.	41.2 $\pm$ 3.7	13.7 $\pm$ 2.2	22.0 $\pm$ 5.5
	6.25	49.4 $\pm$ 3.3	24.7 $\pm$ 2.0	N.D.	0.30 $\pm$ 7.2	3.3 $\pm$ 1.5	8.7 $\pm$ 2.6
IC <sub>50</sub> ( $\mu\text{M}$ )		6.25 $\pm$ 2.9	15.6 $\pm$ 4.2	>100	43.8 $\pm$ 7.3	40.0 $\pm$ 1.7	28.8 $\pm$ 1.9
<b>15</b>	50	100 $\pm$ 7.0	100 $\pm$ 5.6	100 $\pm$ 8.5	100 $\pm$ 6.7	100 $\pm$ 6.5	100 $\pm$ 6.8
	25	100 $\pm$ 5.3	100 $\pm$ 7.2	50.6 $\pm$ 4.7	82.1 $\pm$ 4.9	70.5 $\pm$ 7.3	60.0 $\pm$ 7.2
	12.5	24.9 $\pm$ 4.7	55.0 $\pm$ 1.2	33.5 $\pm$ 3.0	48.1 $\pm$ 8.1	34.8 $\pm$ 3.9	37.8 $\pm$ 4.5
	6.25	20.6 $\pm$ 3.0	27.0 $\pm$ 3.2	22.2 $\pm$ 5.5	36.8 $\pm$ 3.2	23.1 $\pm$ 4.3	23.4 $\pm$ 6.3
	3.125	15.5 $\pm$ 2.1	5.8 $\pm$ 1.8	29.7 $\pm$ 6.3	33.8 $\pm$ 5.0	19.9 $\pm$ 2.9	24.7 $\pm$ 2.9
IC <sub>50</sub> ( $\mu\text{M}$ )		14.3 $\pm$ 2.5	11.3 $\pm$ 3.5	27.3 $\pm$ 5.0	12.3 $\pm$ 4.1	18.3 $\pm$ 2.6	21.3 $\pm$ 3.2

<sup>a</sup> The method is described in the Experimental Section. Each datum represents the mean of three independent experiments. N.D.: not determined.

the two methoxyl signals and the two aromatic protons, indicating that the methoxyl groups are located at C-7 and C-8. Therefore, compound **2** was characterized as 1,2,6-trihydroxy-7,8-dimethoxy-3-methylanthraquinone.

Compound **3** has the molecular formula C<sub>16</sub>H<sub>12</sub>O<sub>6</sub>, confirmed by HREIMS. The <sup>1</sup>H NMR spectrum of **3** shows signals for a methyl group ( $\delta$  2.29), a methoxyl group ( $\delta$  3.80), and a *peri*-hydroxy group ( $\delta$  13.12), an isolated aromatic proton ( $\delta$  7.79), and two meta-coupled protons ( $\delta$  6.56 and 7.06). On acetylation, **3** also yielded a triacetate. In the HMBC spectrum, the C-10 signal at  $\delta$  181.6 was correlated with resonances at  $\delta$  7.79 (H-4) and 7.06 (H-5), the signal at  $\delta$  156.5 (C-2) correlated with the CH<sub>3</sub>-3 ( $\delta$  2.29) and H-4 ( $\delta$  7.79) signals, and the resonance at  $\delta$  147.9 (C-1) correlated with the OCH<sub>3</sub>-1 ( $\delta$  3.80) signal. These data suggested the presence of three hydroxyl groups located at C-2, C-6, and C-8 and the methoxyl group located at C-1. Therefore, compound **3** was identified as 2-hydroxyemodin 1-methyl ether.

All of the isolated compounds were tested against a panel of cancer cell lines according to established protocols.<sup>17</sup> As shown in Table 1, compound **15** displayed cytotoxicity against all six tumor cell lines, while compounds **3** and **10** significantly suppressed the growth of Vero, Wish, Calu-1, Raji, and HeLa tumor cells. All other test compounds had IC<sub>50</sub> values higher than 100  $\mu\text{M}$  against all cell lines. These results suggest that the 1,3,8-trihydroxy for the anthraquinone plays a significant role in the cytotoxic activity.

## Experimental Section

**General Experimental Procedures.** Melting points were determined with a Yanagimoto micro-melting point apparatus and are uncorrected. IR spectra were obtained as KBr pellets on a Nicolet Avatar 320 IR spectrometer. UV spectra were obtained on a Hitachi U-3200 spectrophotometer in MeOH. <sup>1</sup>H, <sup>13</sup>C, and 2D NMR spectra were measured with a Varian Inova-500 spectrometer with deuterated solvent as internal standard. EIMS, HREIMS, and APCIMS were recorded on a Finnigan MAT 95S and Finnigan LCQ spectrometer, respectively.

**Plant Material.** The stems of *Ventilago leiocarpa* were collected at Shihting, Taipei, Taiwan, in October 1999. A voucher specimen (No. 195898) has been deposited in the herbarium of the Department of Botany of the National Taiwan University.

**Extraction and Isolation.** The stems of *V. leiocarpa* (8.5 kg) were extracted with 95% EtOH (50 L  $\times$  3). The ethanolic extracts were combined and concentrated under vacuum to a volume of 1.5 L. The concentrated ethanolic extract was then partitioned successively between H<sub>2</sub>O and EtOAc, followed by *n*-BuOH (each 1 L  $\times$  3). The EtOAc extract (110 g) was subjected to silica gel column chromatography with a gradient of EtOAc in *n*-hexane, and 11 fractions were collected. Fraction 2 (2.6 g) was rechromatographed over silica gel (*n*-hexane) and further purified by preparative TLC (25% benzene/*n*-hexane) to give compounds **1**, **4**, **5**, and **6**. Fractions 4 and 5 were combined (1.7 g) and purified by Sephadex LH-20 (EtOAc) and silica gel column chromatography (12–26  $\mu\text{m}$ , performed at pressure  $\sim$ 10 bar, 2% EtOAc/*n*-hexane) to give **7**, **8**, and **9**. A precipitate from fraction 7 was recrystallized with EtOAc/*n*-hexane to give **10**. The filtrate from fraction 7 was purified on a silica gel column (10% EtOAc/*n*-hexane) and Sephadex LH-20 (MeOH) to yield **10** and **11**. Fraction 10 (6.9 g) gave **2**, **12**, **13**, **14**, and **15** after repeated Sephadex LH-20 (MeOH, acetone) and silica gel column chromatography. Fraction 8 was further purified by silica gel column chromatography (40–63  $\mu\text{m}$ , performed at pressure  $\sim$ 10 bar, 10% EtOAc/benzene) to yield **3**.

**Islandicin 4-methyl ether (1):** red needles (EtOAc/*n*-hexane); mp 184–186 °C; UV (MeOH)  $\lambda_{\text{max}}$  (log  $\epsilon$ ) 441 (3.85), 287 (3.80), 251 (4.12), 227 (4.42) nm; IR (KBr)  $\nu_{\text{max}}$  3400 (OH), 1667, 1620 (C=O) cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  2.41 (3H, s, CH<sub>3</sub>-3), 3.89, (3H, s, OCH<sub>3</sub>-4), 7.18 (1H, s, H-2), 7.27 (1H, d, *J* = 7.5 Hz, H-7), 7.68 (1H, t, *J* = 7.5 Hz, H-6), 7.81 (1H, d, *J* = 7.5 Hz, H-5), 12.06 (OH-8), 12.51 (OH-1); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  17.2 (3-CH<sub>3</sub>), 61.0 (4-OCH<sub>3</sub>), 114.0 (C-9a), 115.5 (C-8a), 120.0 (C-5), 123.6 (C-7, 4a), 126.9 (C-2), 134.8 (C-10a), 137.1 (C-6), 146.6 (C-3), 153.8 (C-4), 159.6 (C-1), 162.0 (C-8), 181.4 (C-10), 192.3 (C-9); HMBC correlations, C-1  $\rightarrow$  OH-1, H-2; C-2  $\rightarrow$  OH-1, 3-CH<sub>3</sub>; C-4  $\rightarrow$  4-OCH<sub>3</sub>, H-2, 3-CH<sub>3</sub>; C-7  $\rightarrow$  OH-8, H-5; C-8  $\rightarrow$  OH-8, H-6; C-8a  $\rightarrow$  OH-8, H-5, H-7; C-9a  $\rightarrow$  OH-1, H-2; C-10  $\rightarrow$  H-5; EIMS *m/z* 284 [M]<sup>+</sup>; HREIMS *m/z* 284.0696 [M]<sup>+</sup> (calcd 284.0685 for C<sub>16</sub>H<sub>12</sub>O<sub>5</sub>).

**Diacetate 1a.** Acetylation of compound **1** (AC<sub>2</sub>O/pyridine) gave **1a**: <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  2.41 (3H, s, CH<sub>3</sub>-3), 2.42, 2.43 (each 3H, s, OAc  $\times$  2), 3.93, (3H, s, OCH<sub>3</sub>-4), 7.23 (1H, s, H-2), 7.35 (1H, d, *J* = 8.0 Hz, H-7), 7.73 (1H, t, *J* = 8.0 Hz, H-6), 8.13 (1H, d, *J* = 8.0 Hz, H-5); EIMS *m/z* 368 [M]<sup>+</sup>, 326 [M - 42]<sup>+</sup>, 284 [M - 42  $\times$  2]<sup>+</sup>.

**1,2,6-Trihydroxy-7,8-dimethoxy-3-methylanthraquinone (2):** yellow needles (EtOAc/*n*-hexane); mp 271–273 °C; UV (MeOH)  $\lambda_{\text{max}}$  (log  $\epsilon$ ) 410 (3.86), 313 (4.14), 283 (4.57), 317 (4.13), 301 (4.17), 225 (4.24) nm; IR (KBr)  $\nu_{\text{max}}$  3453 (OH), 1630 (C=O), 1561, 1456, 1299, 1272 cm<sup>-1</sup>; <sup>1</sup>H NMR (acetone-*d*<sub>6</sub>)  $\delta$  2.35 (3H, s, CH<sub>3</sub>-3), 3.98, 3.99 (each 3H, s, OCH<sub>3</sub>  $\times$  2), 7.55

(1H, s, H-4), 7.57 (1H, s, H-5), 8.92, 9.64 (OH × 2), 13.33 (OH-1); <sup>13</sup>C NMR (acetone-*d*<sub>6</sub>) δ 16.3 (3-CH<sub>3</sub>), 61.5, 61.7 (OCH<sub>3</sub> × 2), 111.9 (C-5), 116.3 (C-9a), 120.2 (C-8a), 122.4 (C-4), 124.3 (C-10a), 131.3 (C-3), 132.9 (C-4a), 147.3 (C-7), 150.4 (C-1), 150.8 (C-2), 156.6 (C-8), 157.4 (C-6), 181.0 (C-10), 188.8 (C-9); HMBC correlations, C-1 → OH-1; C-2 → 3-CH<sub>3</sub>, H-4; C-3 → 3-CH<sub>3</sub>, OH-2; C-4 → 3-CH<sub>3</sub>; C-6 → H-5; C-7 → 7-OCH<sub>3</sub>, H-5; C-8 → 8-OCH<sub>3</sub>; C-8a → H-5; C-9a → OH-1, H-4; C10 → H-4, H-5; EIMS *m/z* 330 [M]<sup>+</sup>; HREIMS *m/z* 330.0736 [M]<sup>+</sup> (calcd 330.0740 for C<sub>17</sub>H<sub>14</sub>O<sub>7</sub>).

**Triacetate 2a.** Acetylation of compound **2** (AC<sub>2</sub>O/pyridine) gave **2a**: <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 2.35 (3H, s, CH<sub>3</sub>-3), 2.38, 2.39, 2.48 (each 3H, s, OAc × 3), 3.94, 4.01, (each 3H, s, OCH<sub>3</sub> × 2), 7.80 (1H, s, H-5), 8.08 (1H, s, H-4); EIMS *m/z* 456 [M]<sup>+</sup>, 414 [M - 42]<sup>+</sup>, 372 [M - 42 × 2]<sup>+</sup>, 330 [M - 42 × 3]<sup>+</sup>.

**2-Hydroxyemodin 1-methyl ether (3):** yellow needles (EtOAc/*n*-hexane); mp 292–294 °C; UV (MeOH) λ<sub>max</sub> (log ε) 393 (3.92), 311 (4.08), 284 (4.42), 228 (4.36) nm; IR (KBr) ν<sub>max</sub> 3422 (OH), 1629 (C=O), 1582, 1455, 1377, 1314, 1267 cm<sup>-1</sup>; <sup>1</sup>H (DMSO-*d*<sub>6</sub>) δ 2.29 (3H, s, CH<sub>3</sub>-3), 3.80, (3H, s, OCH<sub>3</sub>-1), 6.56 (1H, br s, H-7), 7.06 (1H, br s, H-5), 7.79 (1H, s, H-4), 10.30 (OH-2), 11.12 (OH-6), 13.12 (OH-8); <sup>13</sup>C (DMSO-*d*<sub>6</sub>) δ 17.2 (3-CH<sub>3</sub>), 61.9 (1-OCH<sub>3</sub>), 107.8 (C-5), 108.1 (C-7), 111.0 (C-8a), 124.4 (C-9a), 125.6 (C-4a), 126.7 (C-4), 132.6 (C-3), 135.3 (C-10a), 147.9 (C-1), 156.5 (C-2), 164.9 (C-6), 165.3 (C-8), 181.6 (C-10), 187.2 (C-9); HMBC correlations, C-1 → 1-OCH<sub>3</sub>; C-2 → 3-CH<sub>3</sub>, H-4; C-4 → 3-CH<sub>3</sub>; C-5 → H-7; C-8 → OH-8, H-7; C-8a → H-5, H-7, OH-8; C10 → H-4, H-5; APCIMS *m/z* 301 [M + H]<sup>+</sup>; HREIMS *m/z* 300.0652 [M]<sup>+</sup> (calcd 300.0634 for C<sub>16</sub>H<sub>12</sub>O<sub>6</sub>).

**Triacetate 3a.** Acetylation of compound **3** (AC<sub>2</sub>O/pyridine) gave **3a**: <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 2.34 (3H, s, CH<sub>3</sub>-3), 2.37, 2.41, 2.47 (each 3H, s, OAc × 3), 3.88 (3H, s, OCH<sub>3</sub> × 2), 7.98 (1H, s, H-4), 7.94 (1H, br s, H-5), 7.25 (1H, br s, H-7); EIMS *m/z* 426 [M]<sup>+</sup>, 384 [M - 42]<sup>+</sup>, 342 [M - 42 × 2]<sup>+</sup>, 300 [M - 42 × 3]<sup>+</sup>.

**Cell Lines.** The K562, Raji, Vero, Calu-1, HeLa, and Wish cell lines were utilized as target cells in the cytotoxic assay. K562 and Raji cells are erythroleukemia and EBV-transformed B cell lines, respectively (American Type Culture Collection, ATCC, Rockville, MD). They were cultured in RPMI-1640 medium (Hyclone, Logan, UT) containing 10% fetal calf serum (FCS, Gibco, Grand Island, NY), 100 u/mL penicillin, and 100 μg/mL streptomycin. The Vero cell is a green monkey kidney tumor cell line (ATCC, Rockville, MD). The Wish cell is a transformed epithelial cell line, and the Calu-1 cell is a human

lung carcinoma cell line (ATCC, Rockville, MD). The HeLa cell is a human cervical carcinoma cell line (ATCC, Rockville, MD). The Vero, Wish, Calu-1, and HeLa cell lines were cultured in MEM medium containing 10% FCS, 100 μg/mL streptomycin, and 100 u/mL penicillin. These cell lines were cultured at 37 °C in an atmosphere of humidified 5% CO<sub>2</sub>.

**Growth Inhibition Assay.** Growth inhibition was assessed as described previously.<sup>17</sup> Each tumor cell line was cultured with or without various concentrations of compound **3**, **10**, or **15** for 3 days, after which [<sup>3</sup>H]-thymidine was added and incubation continued for 16 h before harvest. Radioactivity was determined by a scintillation counter, and inhibitory activity was calculated as the IC<sub>50</sub> (inhibition of 50% cell proliferation).

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## References and Notes

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